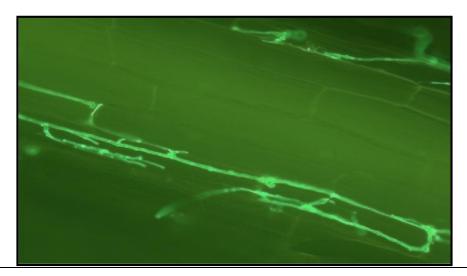


Characterization of phosphate solubilisation and uptake capacity by root fungal endophytes

<u>Diana Rocio Andrade-Linares</u> Benard Ngwene Philipp Franken



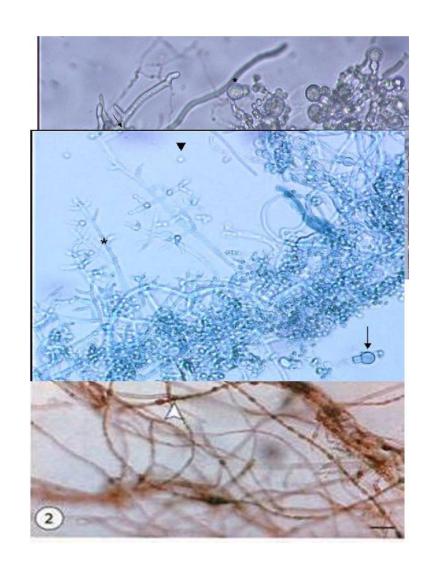
COST FA1103: Endophytes in Biotechnology and Agriculture 14 - 16/11/12, S. Michele all'Adige, Trento, Italy.

Facultative symbiontes in roots

Piriformospora indica

Trichoderma harzianum

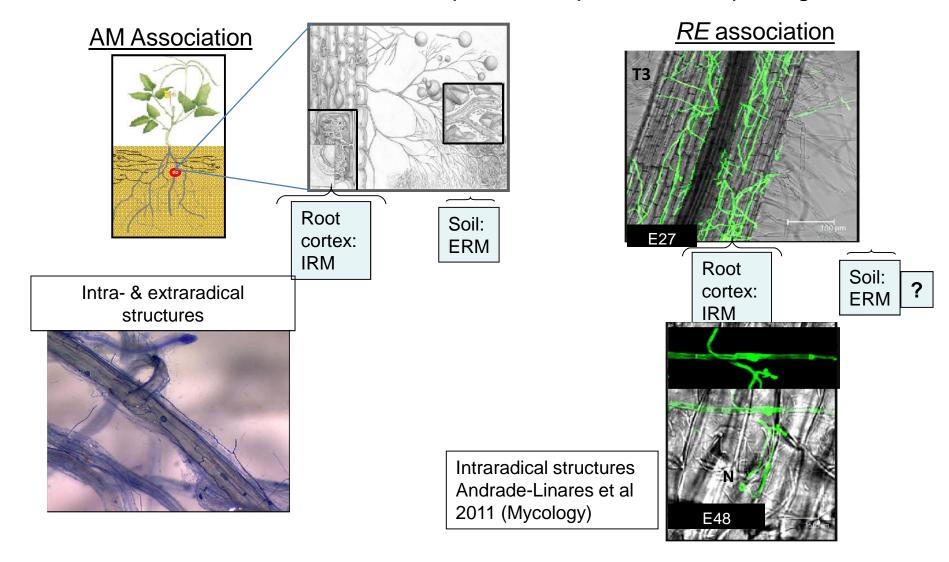
Dark septate endophytes DSE



$\underline{introduction}$

Fungal endophytes

Contribution to mineral element uptake and promotion of plant growth





Fungal endophytes

Research focus

- To evaluate phosphate solubilization of organic and inorganic phosphate sources and the potential production of phosphatases in liquid medium
- To test the capability of *Piriformospora indica* to use organic nitrogen in vitro system
- To evaluate inorganic phosphate transfer, plant growth promotion in tomato plants.
- To evalute ERM growth of *P. indica* into root inaccessible soil compartments
- To analyze the interaction between P. indica and Glomus intraradicis in the symbiosis with tomato plants

General methodology

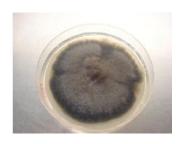
- 1. Phosphate solubilization
 - Lecithin
 - Phytin
 - Pikovskaya medium
 - Rock Phosphate



Agar
Different concentrations

Liquid culture Available Phosphorous and enzimatic activity

DSE48



L. orchidicola 135



P. indica



2. Protease activity in vitro on Casein source

3. Phosphorous uptake in pot experiments

1. Phosphate solubilization



- Lecithin
- Phytin
- Pikovskaya agar
- Phosphate concentrations:
 500, 200, 100, 50, 0 mg/l
- Growth analysis and hydrolisis during 30 days of incubation

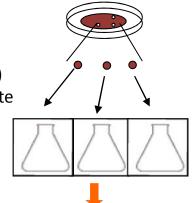
2. Available Phosphorous and enzimatic activity

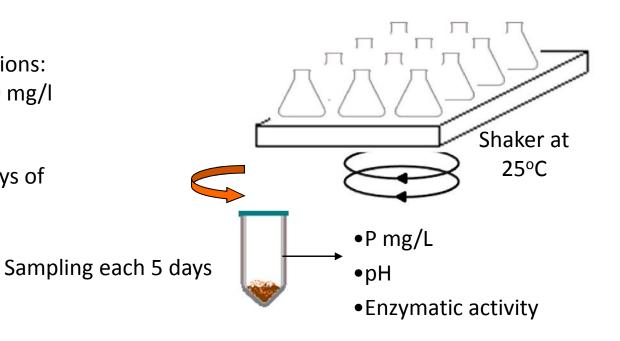
Liquid culture

- KH₂PO₄ (499 mg/L P)
- Tri-calcium phosphate

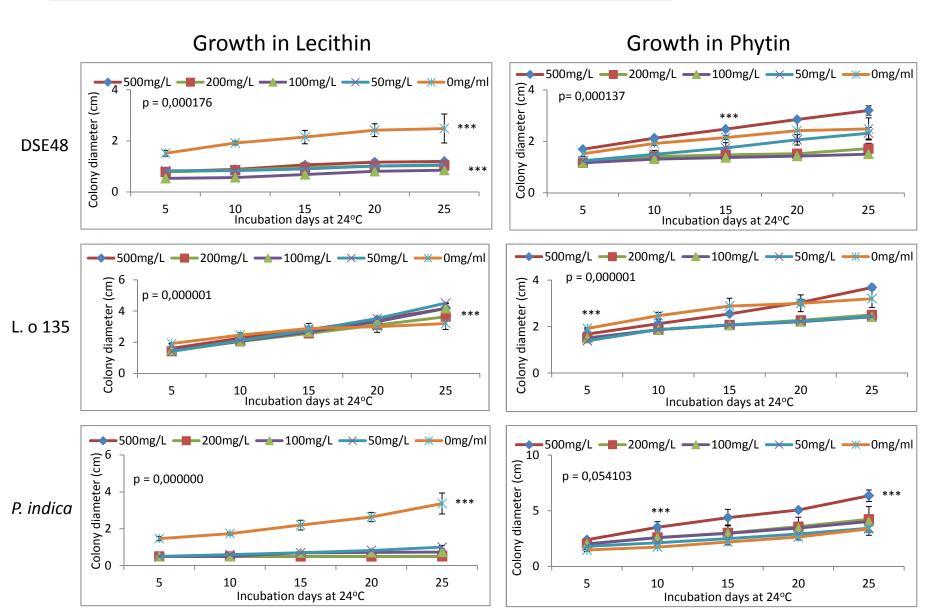
 $(TCP, Ca_3O_8P_2; 0.25\%)$

- Rock Phosphate
 (RP, P₂O₅; 0.347%)
- Lecithin
- Phytin



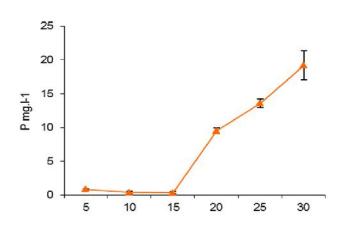


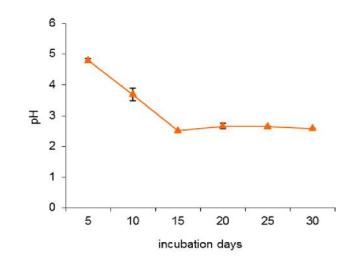
Assimilation and solubilization of different P- sources



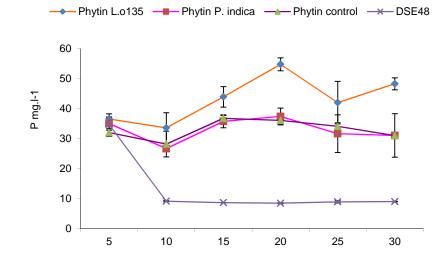
Available Phosphorous and Enzimatic activity

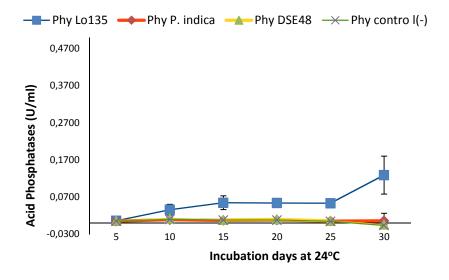
Lecithin 500 mg/L





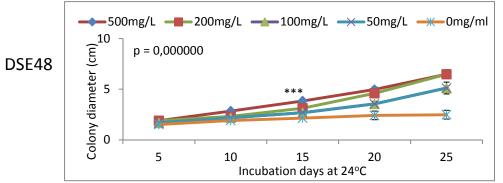
Phytin 150 mg/L

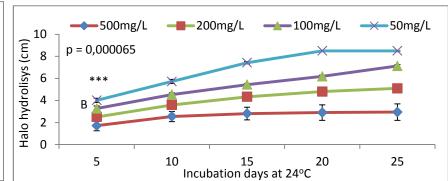


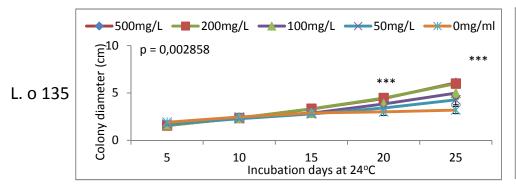


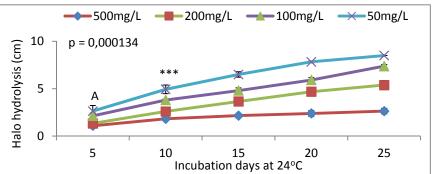
Assimilation and solubilization of different P- sources

Pikovaskaya agar with different concentrations of $Ca_3(PO_4)_2 = (Ca_3-P)$

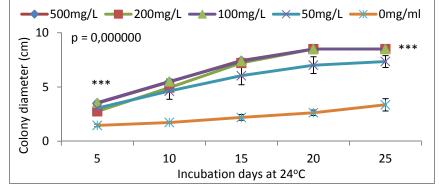






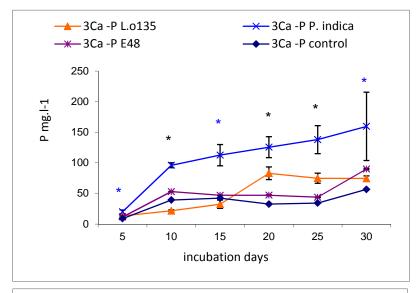


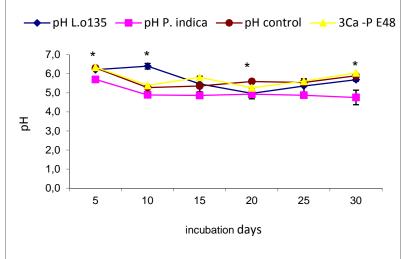




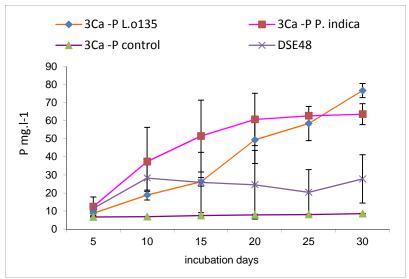
Available Phosphorous and Enzimatic activity

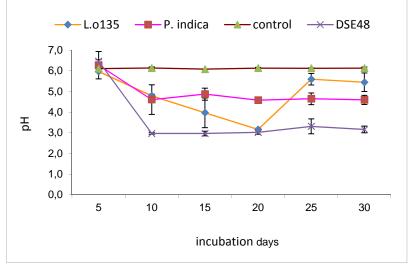
Ca₃-P 500 mg/L





Ca₃-P 150 mg/L

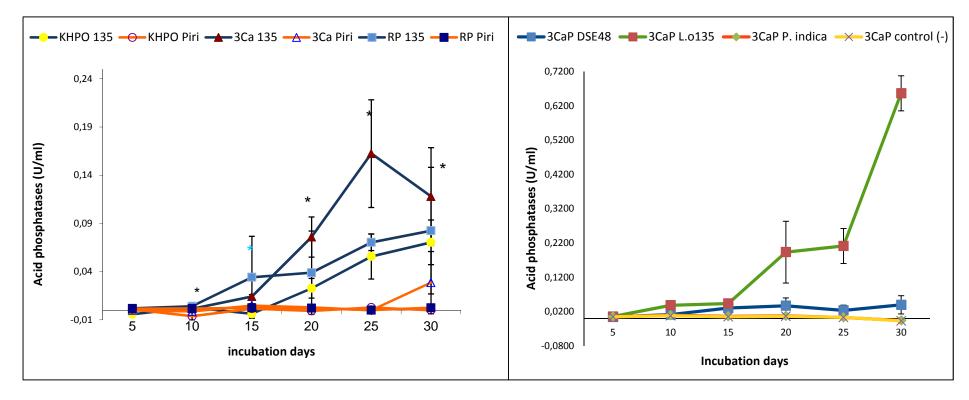




Available Phosphorous and Enzimatic activity

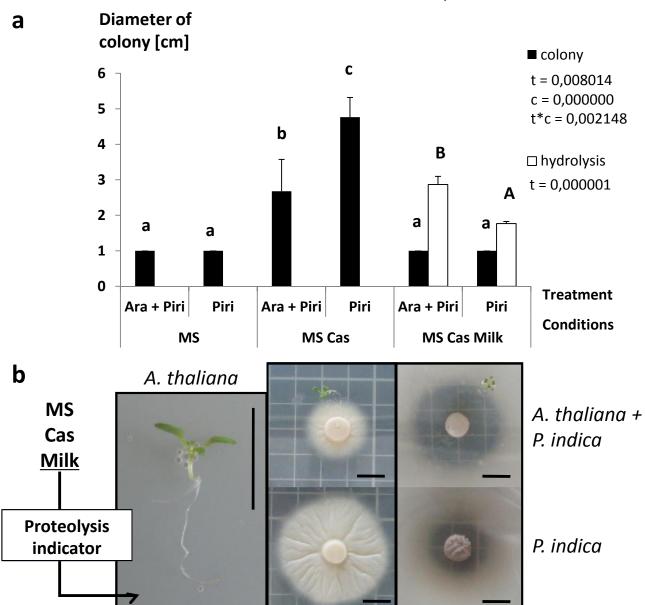
Ca3-P 500 mg/L

Ca3-P 150 mg/L



2. Protease activity

(Brunel, J. et al. 2012)



ntroduction materials & methods results & discussion

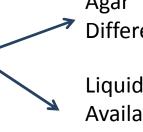


Conclusions

- Lecithin inhibits growth of the DSE48 and *P. indica*, while *L. orchidicola* 135 is able to growth up to a concentration of 500 mg/L, however the release of P_i is low (20mg/L) and after 20 days of incubation.
- The endophytes growth in high concentration of Phytin, specially *P. indica*, however it did not release P_i and not present enzymatic activity in contrast to *L. orchidicola* 135.
- The fungal endophytes grow better at concentrations higher than 100 mg/L in Pikovskaya (Ca₃-P) agar but the solubilization was less evident at 500 mg/L (PO₄)_{2.}
- P. indica does not produce hydrolysis halo in comparison to the DSE in Ca₃-P but it is able to release P_i in liquid medium and seems to be more efficient at higher concentration (500 mg/L) in contrast to L. orchidicola 135. However P. indica does not have extracellular phosphatase activity.
- *P. indica* does produce hydrolysis halo in casein agar which indicate the production of proteases which seems to be regulated by the host plant.

General methodology

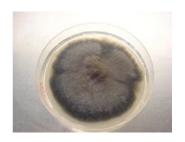
- 1. Phosphate solubilization
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Agar
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DSE48



L. orchidicola 135



P. indica



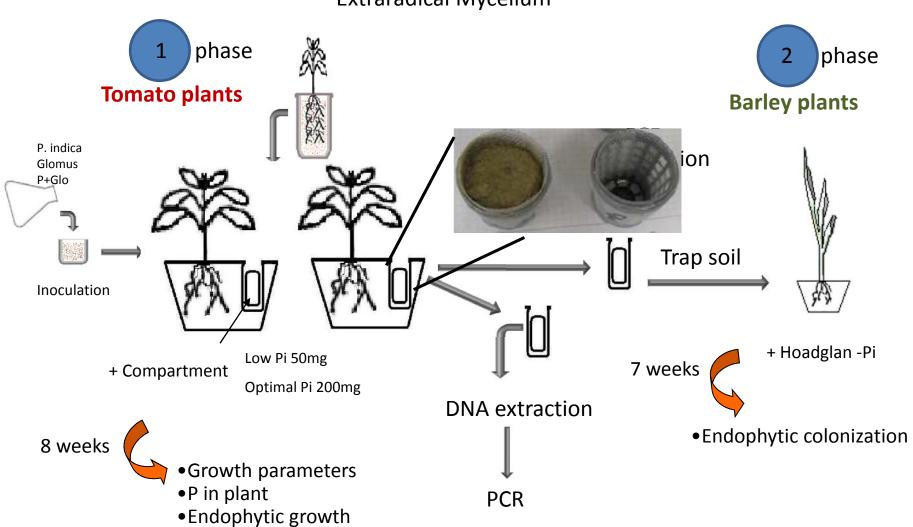
2. Protease activity in vitro on Casein source

3. Phosphorous uptake in pot experiments

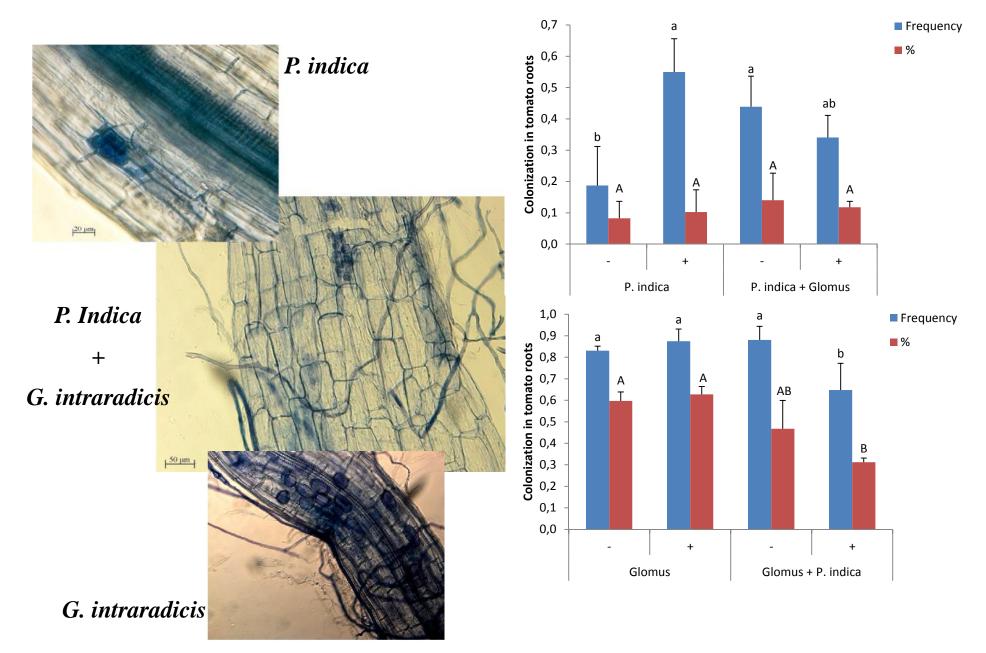
3. Phosphorous uptake: Pi content in plant

Root colonization

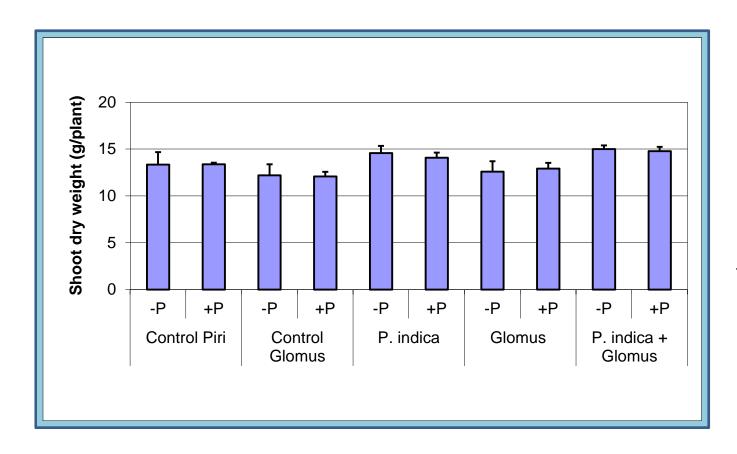
Extraradical Mycelium



Tomato colonized roots (frequency and intensity)

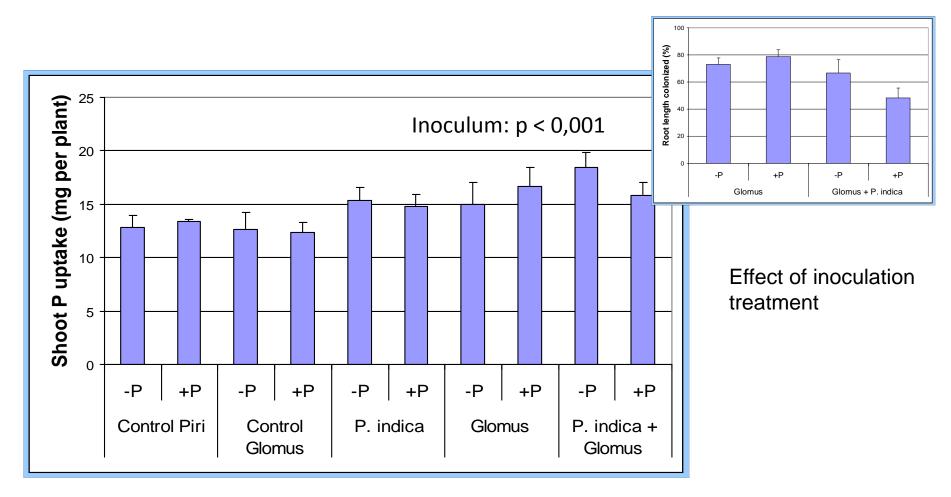


Shoot dry weight after harvest (g per plant)



Effect of inoculation treatment

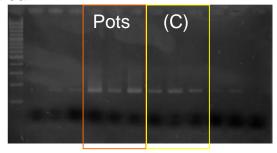
Shoot P uptake after harvest (mg per plant)



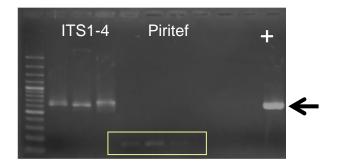
Detection of extraradical mycelium of P. indica

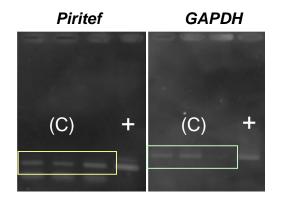
Soil of compartments

P. Indica in:



Soil for trap pots with barley



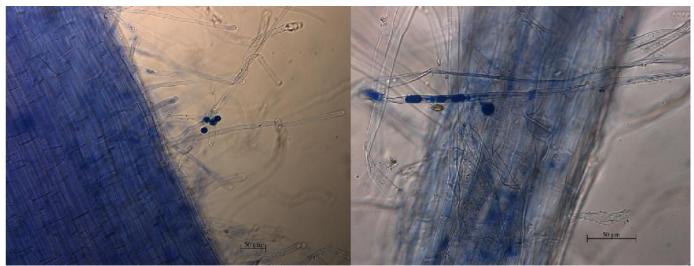


Primer pair for the translation elongation factor 1- gene (TEF): *Piritef*

Primer pair for the glyceraldehyde-3-phosphate dehydrogenase gen (*GAPDH*)

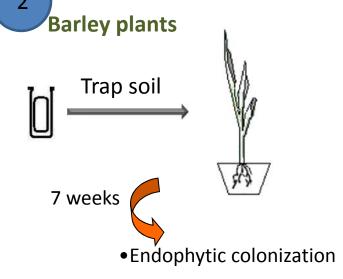
P. indica and G. intraradicis in Compartment (C)

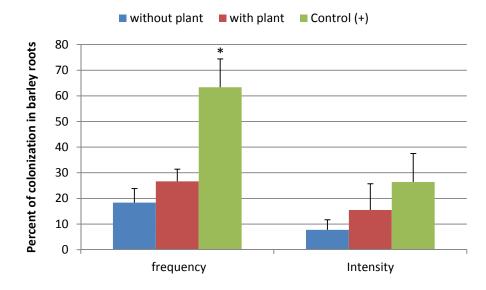
Colonization of trap plant roots



P. indica spores and hyphae attaching root hair of barley plant

P. indica spores inside root hair of barley plant





ntroduction materials & methods results & discussion



Conclusions

- Tomato root plants were colonized at the same time by *P. indica* and *G. intraradicis*. This colonization seems to be lower for *P. indica* than for *G. intraradicis*.
- However the AMF colonization was negatively affected in the combination treatment (Piri + Glomus) at high concentration of P_i in the compartment.
- The inoculation with the endophyte *P. indica* showed a significant effect on plant dry weight as well as on P uptake and this effect is higher with the presence of the AMF in roots.
- *P. indica* is able to produce ERM which was detected by PCR in soil compartments and was also recovered in the roots of trap plants.

- •Phosphorous and nitrogen uptake from organic sources and interaction with AMF.
- •Quantification of extraradical mycelium and root colonization at saprophytic and endophytec stages in soil.
- Regulation of phosphate transportes
- •Influence of endohyphal bacteria on fungal and plant nutrition.

